

PRACA ORYGINALNA

# Effect of chlorpheniramine and cimetidine, a histamine H<sub>1</sub> and H<sub>2</sub> antagonist on (<sup>3</sup>H)glucose uptake in the brain of adult rats lesioned with 5,7-dihydroxytryptamine as neonates

Wpływ chlorfeniraminy i cymetydyny, antagonistów receptorów histaminowych H<sub>1</sub> i H<sub>2</sub> na wychwyt (<sup>3</sup>H)glukozy w mózgu dorosłych szczurów po podaniu noworodkom 5,7-dihydroksytryptaminy

Jadwiga Jośko<sup>1</sup>, Jacek Drab<sup>1</sup>, Przemysław Nowak<sup>2</sup>, Ryszard Szkilnik<sup>2</sup>,  
Dariusz Boroń<sup>3</sup>, Marta Elwart<sup>2</sup>, Janusz Konecki<sup>3</sup>, Halina Brus<sup>3</sup>, Ryszard Brus<sup>2,4</sup>

## A B S T R A C T

### BACKGROUND

The aim of the study was to examine effect of chlorpheniramine (histamine H<sub>1</sub> receptor antagonist) and cimetidine (histamine H<sub>2</sub> receptor antagonist) on (<sup>3</sup>H)glucose uptake in the brain of adult rats lesioned with 5,7-dihydroxytryptamine (neurotoxin for the central serotonergic system) as neonates.

### MATERIAL AND METHODS

Male 3-days old Wistar rats were injected with serotonergic neurons neurotoxin 5,7-dihydroxytryptamine, 75 µg *icv*. Control rats were injected with saline 10 µg *icv*. At 8 weeks level of 5-HT and 5-HIAA was estimated in the striatum, frontal cortex and hippocampus of the brain. Other 8 weeks old animals of control and 5,7-DHT lesioned as neonates were injected with S(+)-chlorpheniramine (H<sub>1</sub> receptor antagonist) 10.0 mg/kg *ip* or with cimetidine (H<sub>2</sub> receptor antagonist) 5.0 mg/kg *ip*. Control rats were injected with saline 1.0 ml/kg *ip*. 60 minutes later 6-(<sup>3</sup>H)-D-glucose was applied in a dose of 500 µCi/kg *ip* and 15 minutes later all rats were decapitated and their brains were excised, placed on the ice and sample of frontal cortex, striatum, hippocampus, thalamus with hypothalamus, pons and cerebellum were separated and weighted. Then in the examined tissues radioactivity was measured in liquid scintillation counter and expressed in DPM/100 mg of wet tissue.

<sup>1</sup>Chair and Department of Medicine and Environmental Epidemiology,

<sup>2</sup>Department of Pharmacology,

<sup>3</sup>Department of Histology and Embriology  
School of Medicine with the Division  
of Dentistry in Zabrze, Medical University  
of Silesia in Katowice

<sup>4</sup>High School of Strategic Planning  
in Dąbrowa Górnica

### ADRES

#### DO KORESPONDENCJI:

Prof. dr hab. n. med. Jadwiga Jośko

Chair and Department of Medicine  
and Environmental Epidemiology  
School of Medicine with the Division

of Dentistry in Zabrze  
Medical University of Silesia in Katowice  
ul. Jordana 19  
41-808 Zabrze, Poland  
tel./fax 32 27 22 847  
e-mail: jjosko@sum.edu.pl

**RESULTS**

5,7-DHT decreased significantly the level of 5-HT and 5-HIAA in all examined tissues in the brain of adult rats. In rats neonatally lesioned with 5,7-DHT radioactivity significantly increased as compare to the control. Chlorpheniramine prevent significantly that effect in the frontal cortex and cimetidine in the frontal cortex, hippocampus and cerebellum.

**CONCLUSION**

From above we conclude that in the brain of mammals the metabolic link between histaminergic and serotonergic system exist in regulation of energetic processes connected with glucose metabolism.

**KEY WORDS**

5,7-DHT, 5-HT, 5-HIAA, chlorpheniramine, cimetidine, 6-<sup>3</sup>H-D-glucose, brain, rats

**S T R E S Z C Z E N I E****WSTĘP**

Celem pracy było zbadanie wpływu chlorfeniraminy, antagonisty receptora histaminowego H<sub>1</sub> i cymetydyny, antagonisty receptora histaminowego H<sub>2</sub> na wychwyt (<sup>3</sup>H)glukozy w mózgu dorosłych szczerów z leżą (zniszczanie) ośrodkowego układu serotonergicznego wywołaną podaniem noworodkom neurotoksyny 5,7-dihydroksytryptaminy.

**MATERIAŁ I METODY**

Trzydniowe noworodki płci męskiej szczebu Wistar otrzymały do bocznej komory mózgu (*icv*) 75 µg 5,7-dihydroksytryptaminy (5,7-DHT), neurotoksynę układu serotonergicznego. Zwierzęta kontrolne otrzymały *icv* 10 µl 0,9% roztworu NaCl. Po osiągnięciu 8 tygodni życia zwierzęta dekapitowano i w korze czołowej, prążkowiu oraz zakręcie hipokampa oznaczono zawartość 5-HT i 5-HIAA metodą HPLC/ED. Osobnej grupie badanej i kontrolnej podano S(+)-chlorfeniraminę 10,0 mg/kg *ip* (antagonista receptora histaminowego H<sub>1</sub>) lub cymetydynę 5,0 mg/kg *ip* (antagonista receptora histaminowego H<sub>2</sub>). Zwierzęta kontrolne obu grup otrzymały 0,9% roztwór NaCl 1,0 ml/kg *ip*. Po 60 minutach wszystkie szczerury otrzymały 6-(<sup>3</sup>H)-D-glukozę 500 µCi/kg *ip*. Po dalszych 15 minutach zwierzęta dekapitowano, wyjmowano z czaszki mózg, separowano z niego korę czołową, prążkowie, hipokamp, wzgórze z podwzgórzem, most i mózdzek, w których oznaczono radioaktywność przy użyciu licznika scyntylacyjnego. Wyniki wyrażono w DPM (Desintegrations Per Minute) na 100 mg świeżej tkanki.

**WYNIKI**

5,7-DHT podany noworodkom znamienne obniżył zawartość 5-HT i 5-HIAA w badanych fragmentach mózgu dorosłych szczerów. U zwierząt z leżą ośrodkowego układu serotonergicznego we wszystkich badanych częściach mózgu wykazano znamienne wzrost wychwytu (<sup>3</sup>H)glukozy. Badani antagoniści receptorów histaminowych nie wpływali na wychwyt (<sup>3</sup>H)glukozy w mózgu zwierząt grupy kontrolnej, natomiast chlorfeniramina zapobiegała wychwytowi glukozy tylko w korze mózgowej, a cymetydyna w korze mózgowej, hipokampie i mózdziku zwierząt z leżą ośrodkowego układu serotonergicznego wywołaną podaniem noworodkom 5,7-DHT.

**WNIOSKI**

Wyniki wskazują na metaboliczne powiązania w mózgu ssaków między układem serotonergicznym i histaminergicznym.

**SŁOWA KLUCZOWE**

5,7-DHT, 5-HT, 5-HIAA, chlorfeniramina, cymetydyna, 6-(<sup>3</sup>H)-D-glukosa, mózg, szczerury

## INTRODUCTION

Glucose is the main energy source in mammalian brain. Many factors and agents can influence and modify glucose uptake in brain. We showed that 6-hydroxydopamine (6-OHDA), a neurotoxin for the central dopaminergic system [1] applied to newborn rats in a dose of 134 µg *icv* reduced [<sup>3</sup>H]glucose uptake in the brain of adult rats [2]. In those animals an increase of serotonergic and histaminergic activity was observed in brain [3–5]. Beside we founded that in 5,7-dihydroxytryptamine (5,7-DHT) lesioned rats as neonates increase of (<sup>3</sup>H)glucose uptake in the brain of adult rats, and thioperamide, a central histamine H<sub>3</sub> receptor antagonist prevents this effect [6]. Therefore, the aim of the present study was to examine the effect of 5,7-DHT, a neurotoxin for the central serotonergic system applied to newborn rats on (<sup>3</sup>H)glucose uptake in the brain of adult rats, and effect of chlorpheniramine and cimetidine a histamine receptor H<sub>1</sub> and H<sub>2</sub> antagonist on it.

## MATERIALS AND METHODS

At 3<sup>rd</sup> day after birth male Wistar rat pups were pretreated with 5,7-DHT 75 µg introcerebroventricularly (*icv*), (base form, half on each side with 0.1% ascorbic acid – Sigma MO, USA) or saline with 0.1% ascorbic acid vehicle (5 µl on each side). For details see Brus et al. [4]. Rat's pups remained with their mothers until 21<sup>st</sup> day, then were separated to individual cages. All rats were kept under a 12 h light: 12 h dark cycle in a well ventilated room, at 22 ± 1°C, with free access to food and tap water. This study was approved and controlled by the Local Ethics Committee for Animals at the Medical University of Silesia (permission no 4/2008 issued on 08.01.2008).

When rats attained of 8 weeks biochemical studies were performed. Rats of both groups were scarified by decapitation and the brains were immediately excised and placed on the ice. The corpus striatum, frontal cortex and gyrus hippocampus were separated at temperature of 0°C, placed on dry ice, weighed and stored at -70°C pending of analysis of 5-hydroxytryptamine (5-HT) and 5-hydroxyindolacetic acid (5-HIAA) by an HPLC/ED technique [7]. Results were expressed as ng/g of wet tissue.

Adult rats of both groups (control and neonatally lesioned with 5,7-DHT) were injected with saline vehicle or S(+)-chlorpheniramine (histamine H<sub>1</sub> receptor antagonist) 10.0 mg/kg *ip* (Sigma MO, USA) or with cimetidine (histamine H<sub>2</sub> receptor antagonist) 5.0 mg/kg *ip* (Polfa, Poland). Then 60 minutes later all rats were injected with 6-(<sup>3</sup>H)-D-glucose (Amersham Radiochemicals, Pittsburgh, PA, USA) in a dose of 500 µCi/kg *ip*. 15 minutes later rats were decapitated without anaesthesia. Their brains were immediately excised and placed on ice, for dissection of samples (50–100 mg) of frontal cortex, striatum, hippocampus, thalamus with hypothalamus, pons, and cerebellum. Individual tissues were weighed and placed in 20 ml scintillation vials containing 1 ml of Soluene-350 (Packard Inc., Downers Grove, IL, USA). Each vial was then tightly sealed and incubated at 37°C for 48 h, for solubilization of tissue. Scintillation cocktail Dimilune-30 (Packard Inc., Downers Grove, IL, USA) 10 ml then added and vials were briefly vortexed and placed in a scintillation counter (DSA 14091, Wallac, Finland). Measurement of radioactivity was performed twice for two minutes and the mean was taken. Finally, the mean ± SEM DPM (desintegrations per minute) per 100 mg of wet tissue was calculated for each tissue and group.

Data were analyzed by two-way ANOVA and the post-ANOVA test of Neuman-Kuels. Differences in p values of < 0.05 were considered significant.

## RESULTS

5,7-DHT injected to newborn rats decreased 5-HT and 5-HIAA level in significant way in all examined parts of the brain of adult rats (tab. I).

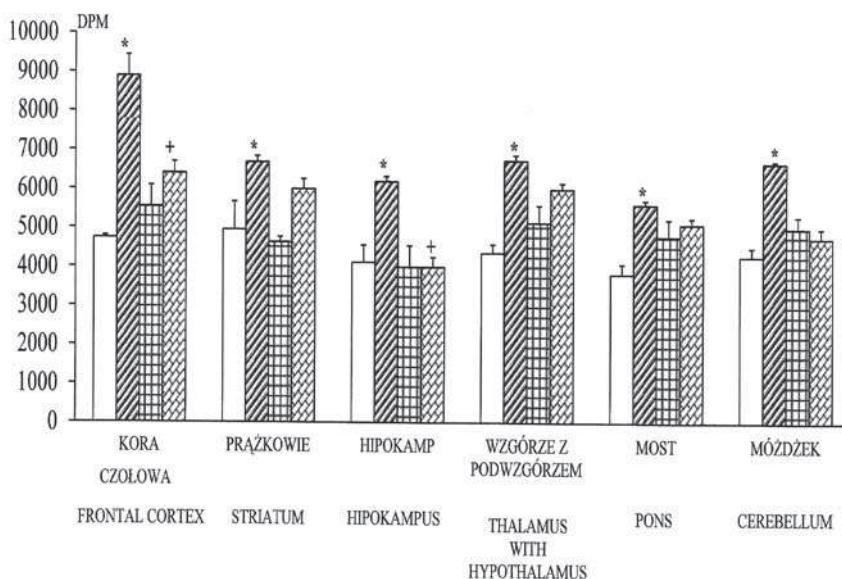
5,7-DHT applied to newborn rats significantly increased (<sup>3</sup>H)glucose uptake in all examined parts of the brain of adult rats as compare to the control group (fig. 1, 2). Chlorpheniramine 10.0 mg/kg *ip* and cimetidine 5.0 mg/kg *ip* did not change (<sup>3</sup>H)glucose uptake in the intact group of rats versus respective control and prevented (<sup>3</sup>H)glucose uptake in the frontal cortex of 5,7-DHT neonatally lesioned adult rats (fig. 1). Cimetidine prevented (<sup>3</sup>H)glucose uptake in the frontal cortex, hippocampus and cerebellum versus respective control (5,7-DHT lesioned rats) (fig. 2).

**Table I.** Effect of 5,7-dihydroxytryptamine 75 µg icv applied to newborn rats on 5-HT and 5-HIAA level in brain of adult rats ( $X \pm SEM$ ; n = 5–8)

**Tabela I.** Wpływ podania dokomorowo 75 µg 5,7-dihydroksytryptaminy noworodkom szczurzym na stężenie 5-HT i 5-HIAA w mózgu dorosłych szczurów ( $X \pm SEM$ ; n = 5–8)

Part of the brain	5-hydroxytryptamine ng/g of wet tissue		5-hydroxyindoleacetic acid ng/g of wet tissue	
	saline 10 µl icv	5,7-dht 75 µg icv	saline 10 µl icv	5,7-dht 75 µg icv
Striatum	562 ± 42	62* ± 21	307 ± 55	96* ± 22
Frontal cortex	244 ± 19	22* ± 9	202 ± 19	35* ± 13
Hippocampus	255 ± 21	43* ± 14	221 ± 16	42* ± 13

\*p < 0.05 versus: saline



**Fig. 1.** Effect of S(+)chlorpheniramine on ( $^3$ H)glucose uptake in the brain of adult rats lesioned with 5,7-dihydroxytryptamine as neonates ( $X \pm SEM$ ; n = 5).

- 0.9% NaCl 10 ml icv as neonates; 0.9% NaCl 1.0 ml/kg ip as adult; ( $^3$ H)glucose 500 µg/kg icv
- 5,7-Dihydroxytryptamine 75 µg icv as neonates; 0.9% NaCl 1.0 ml/kg ip as adult; ( $^3$ H)glucose 500 µCi/kg ip as adult
- 0.9% NaCl 10 µl icv as neonates; S(+)Chlorpheniramine 10.0 mg/kg ip as adult; ( $^3$ H)glucose 500 µCi/kg ip as adult
- 5,7-Dihydroxytryptamine 75 µg icv as neonates; 0.9% NaCl 1.0 ml/kg ip as adult; ( $^3$ H)glucose 500 µCi/kg ip as adult

Explanation:

. p < 0.05 versus to the 0.9% NaCl ip

+\_p < 0.05 versus to the 5,7-DHT icv

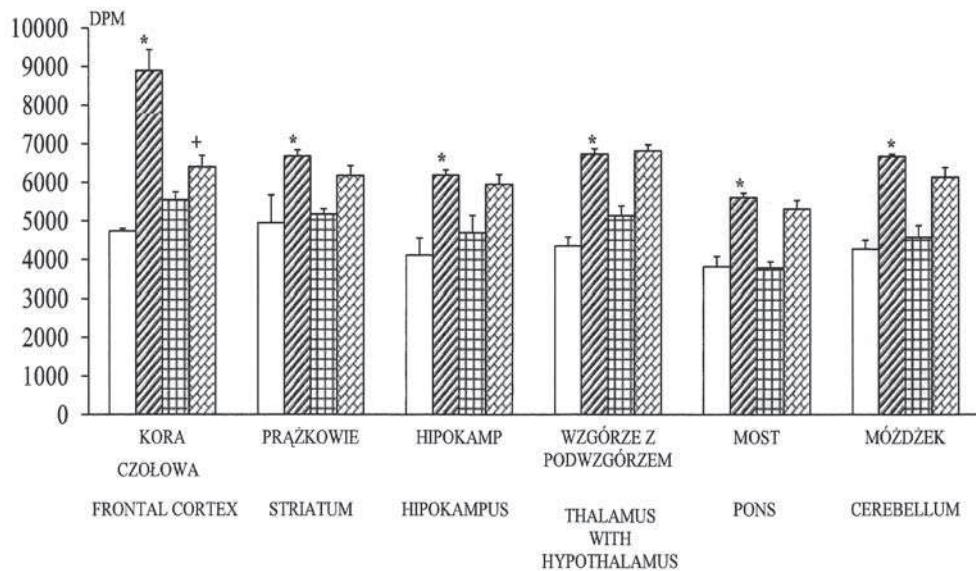
**Ryc. 1.** Wpływ S(+)chlorfeniraminy na wychwyt ( $^3$ H)glukozy w mózgu dorosłych szczurów, u których w okresie noworodkowym dokonano uszkodzenia mózgu 5,7-dihydroksytryptaminy ( $X \pm SEM$ ; n = 5).

- 10 ml 0,9% NaCl podane dokomorowo noworodkom; 1,0 ml/kg 0,9% NaCl podane dootrzewnowo dorosły; 500 µg/kg ( $^3$ H)glukozy podane dokomorowo
- 75 µg 5,7-Dihydroxytryptaminy podane dokomorowo noworodkom; 1,0 ml/kg 0,9% NaCl podane dootrzewnowo dorosły; 500 µCi/kg ( $^3$ H)glukozy podane dootrzewnowo dorosły
- 10 µl 0,9% NaCl podane dokomorowo noworodkom; 10,0 mg/kg S(+)Chlorfeniraminy podane dootrzewnowo dorosły; 500 µCi/kg ( $^3$ H)glukozy podane dootrzewnowo dorosły
- 75 µg 5,7-Dihydroxytryptaminy podane dokomorowo noworodkom; 1,0 ml/kg 0,9% NaCl podane dootrzewnowo dorosły; 500 µCi/kg ( $^3$ H)glukozy podane dootrzewnowo dorosły

Objaśnienie:

. p < 0,05 w porównaniu do 0,9% NaCl podanego dootrzewnowo

+\_p < 0,05 w porównaniu do 5,7-DHT podanego dokomorowo



**Fig. 2.** Effect of cimetidine on (<sup>3</sup>H)glucose uptake in the brain of adult rats lesioned with 5,7-dihydroxytyramine as neonates (X ± SEM; n = 5).

- 0.9% NaCl 10 ml *icv* as neonates; 0.9% NaCl 1.0 ml/kg *ip* as adult; (<sup>3</sup>H)glucose 500 µCi/kg *ip* as adult
- ▨ 5.7-Dihydroxytryptamine 75 µg *icv* as neonates, 0.9% NaCl 1.0 ml/kg *ip* as adult; (<sup>3</sup>H)glucose 500 µCi/kg *ip* as adult
- ▨ 0.9% NaCl 10 µl *icv* as neonates; Cimetidine 5.0 mg/kg *ip* as adult; (<sup>3</sup>H)glucose 500 µCi/kg *ip* as adult
- ▨ 5.7-Dihydroxytryptamine 75 µg *icv* as neonates; Cimetidine 5.0 mg/kg *ip* as adult; (<sup>3</sup>H)glucose 500 µCi/kg *ip* as adult

Explanation:

. p < 0.05 versus to the 0.9% NaCl *ip*

+ p < 0.05 versus to the 5,7-DHT *icv*

**Ryc. 2.** Wpływ cymetydyny na wychwyt (<sup>3</sup>H)glukozy w mózgu dorosłych szczurów, u których w okresie noworodkowym dokonano uszkożenia mózgu 5,7-dihydroksytyraminą (X ± SEM; n = 5).

- 10 ml 0,9% NaCl podane dokomorowo noworodkom; 1,0 ml/kg 0,9% NaCl podane dootrzewnowo dorosłym; 500 µCi/kg (<sup>3</sup>H)glukozy podane dokomorowo
- ▨ 75 µg 5,7-Dihydroksytryptaminy podane dokomorowo noworodkom; 1,0 ml/kg 0,9% NaCl podane dootrzewnowo dorosłym; 500 µCi/kg (<sup>3</sup>H)glukozy podane dootrzewnowo dorosłym
- ▨ 10 µl 0,9% NaCl podane dokomorowo noworodkom; 5,0 mg/kg Cymetydyny podane dootrzewnowo dorosłym; 500 µCi/kg (<sup>3</sup>H)glukozy podane dootrzewnowo dorosłym
- ▨ 75 µg 5,7-Dihydroksytryptaminy podane dokomorowo noworodkom; 5,0 mg/kg Cymetydyny podane dootrzewnowo dorosłym; 500 µCi/kg (<sup>3</sup>H)glukozy podane dootrzewnowo dorosłym

Objaśnienie:

. p < 0,05 w porównaniu do 0,9% NaCl podanego dootrzewnowo

+ p < 0,05 w porównaniu do 5,7-DHT podanego dokomorowo

## DISCUSSION

5,7-DHT is a selective neurotoxin for the central serotonergic system. Applied to newborn rats in a dose of 75 µg *icv* decreased 5-HT and its metabolite – 5-HIAA content in the brain permanently, for whole life of animals. 5,7-DHT after applying to the brain (*icv*) is uptake via active transport like original neurotransmitter (5-HT) to the serotonergic neurons where is metabolized to the very active neurotoxic derivatives induced neurons damage [8]. Damage

central serotonergic system with 5,7-DHT caused changes activity of the other systems, mostly dopaminergic one [4,9].

There are some scarce data concern central serotonergic activity on glucose metabolism in the mammalians brains. Darves and Gudelski [10] presented that activation of the central serotonin 5-HT<sub>2</sub> receptor using respective agonist (DOI) induces glycogenolysis in rat brain. Others presented that quipazine, a 5- HT<sub>3</sub> receptor agonist increased glucose metabolism in the rat's brain measured by quantitative autoradiographic (<sup>14</sup>C)deoxyglucose technique

[11]. Also m-chlorphenylpiperazine (mCPP), a 5-HT<sub>2C</sub> central receptor agonist increased glucose utilization in the brains of healthy volunteers examined by 2-(<sup>18</sup>F)-2-deoxy-D-glucose positron emission tomography [12]. Presented finding that lesion of the central serotonergic system as neonates increased glucose uptake in the adult rats' brain seems to be the first one.

Histamine (H) is a neurotransmitter with wide distribution in human and mammalian's brain [13]. The cell bodies of histaminergic neurons are located mostly in the *tubero- and mamillary nucleus* of poster hypothalamus [14]. Neurons provide histaminergic fibres to the all areas of the mammalian's brain. Since the time when it was recognized that classic H<sub>1</sub> receptor antagonists used as antiallergic drugs exert significant sedative effect, three types of H receptors were discovered in the brain of mammals that is H<sub>1</sub>, H<sub>2</sub> and H<sub>3</sub> [14,15]. The first two are post-synaptic and H<sub>3</sub> seems to be presynaptic one [16,17]. All of them, but mostly H<sub>3</sub> modulate H release from histaminergic neurons in the brain. Thioperamide is an agent which blocks central H<sub>3</sub> receptor and intensified H release from histaminergic neurons to synaptic cleft [14,18,19]. The H<sub>3</sub> central receptors seem to be a main target in alternative treatment of motor dysfunction in Parkinson's disease.

There are few data only concern effect of the central histaminergic system activity on glucose metabolism in the mammalian brains. Injection of H *icv* to the brain of rats induced hyperglycemic response [20]. Ohters founded that pyrilamine and ranitidine, a H<sub>1</sub> and H<sub>2</sub> receptor antagonists respectively, induced hyperglycemia in anesthetized rats [21]. In the next studies it was founded that metaprine a histamine N-methyltransferase inhibitor, elevated plasma glucose concentration in rats [22]. Others presented that 2-deoxy-D-glucose applied to the brain (*icv*) increased glucose plasma level and pretreatment of animals with cimetidine

attenuate observed effect [23]. In our previous study we presented that thioperamide, a histamine H<sub>3</sub> receptor antagonist prevented (<sup>3</sup>H)glucose uptake in all examined parts of the brain of rats neonatally lesioned rats with 5,7-dihydroxytryptamine as neonates [6].

It is difficult to explain inhibitory effect chlorpheniramine and cimetidine a H<sub>1</sub> and H<sub>2</sub> receptor antagonists on the increased (<sup>3</sup>H)glucose uptake in the brain of rats with lesioned central serotonergic system as neonates. Previously we founded that the changed activity of the central dopaminergic system by injection of the neurotoxin 6-hydroxydopamine (6-OHDA) *icv* or a dopamine receptor agonists such as SKF 38393 (D<sub>1</sub> receptor agonist) or quinpirole (D<sub>2</sub>/D<sub>3</sub> receptor agonist) influenced (<sup>3</sup>H)glucose uptake in rats as compare to the control [2,24].

It must be added that 6-OHDA, a neurotoxin of the central dopaminergic system, injected *icv* on the 3<sup>rd</sup> day of life of newborn rats increased histamine level in the brain of adult animals [6]. On the other hand neonatal lesion of the central serotonergic system with 5,7-DHT decreased histamine level in the brain of adult rats [6]. Beside, thioperamide histamine H<sub>3</sub> receptor antagonist diminished reactivity of the central dopamine receptors in 6-OHDA neonatally lesioned rats and in opposite intensified it in 5,7-DHT lesioned as neonates adult rats [25,26]. From above we concluded and confirmed that in the brain of mammals the functional link between histaminergic and serotonergic system exist.

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## EFFECT OF CHLORPHENIRAMINE AND CIMETIDINE

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